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A New Biphenyl Neolignan from Leaves of *Patrinia villosa* (Thunb.) Juss.

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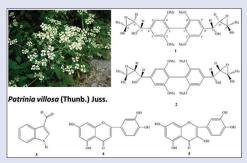
ABSTRACT

Results: One new stereoisomer of biphenylneolignan with four known compounds was isolated from the leaves of Patrinia villosa Juss. **Methods:** The structure of the new compound was elucidated as 2,6,2',6'-tetramethoxy-4,4'-bis (1,2-trans-2,3-epoxy-1-hydroxypropyl) biphenyl (1) on the basis of spectroscopic analysis and comparison with literature data. The four known compounds were identified as 2,6,2',6'-tetramethoxy-4,4'-bis(1,2-cis-2,3-epoxy-1-hydroxypropyl)biphenyl (2), 1H-indole-3-carbaldehydidal (3), luteolin (4) and quercetin(5) by comparison of their spectral data with the reported data, respectively. **Conclusions:** Compound 1 is a new biphenylneolignan, compound 2 and 3 were isolated for the first time from the plant.

Key words: Biphenyl neolignan, coupling constant, nuclear magnetic resonance, *Patrinia villosa* (Thunb.) Juss., structure

SUMMARY

One new stereoisomer of biphenylneolignan named 2,6,2',6'-tetramethoxy-4,4'-bis (1,2-trans-2,3-epoxy-1-hydroxypropyl) biphenyl with four known compounds was isolated from the leaves of Patrinia villosa Juss.



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INTRODUCTION

Patrinia, which belongs to the family of Valerianaceae, is a widely distributed plant grown in East Asia and North America. The genus included more than 20 species, 10 of which growing in China.^[1] Usually, Patrinia species are used as leaves and vegetables in some areas of China, and research also revealed its leaves with pharmacological properties, especially the species of Patrinia villosa (Thunb.) Juss. The P. villosa (Thunb.) Juss., is an important ancient herbal medicine widely used for more than 2000 years from Shen Nong Ben Cao Jing, a famous ancient Chinese medicinal literary. It has been used in traditional Chinese medicine for an inflammation, wound healing, and abdominal pain. The previous research on the chemical constituents of P. villosa (Thunb.) Juss., have revealed that it contains several compound classes. Pentacyclic triterpenoids, iridoids, and flavonoids are the dominant bioactive constituents in the leaves of P. villosa Juss., which displayed potential ability of anti-tumor and anti-inflammatory. [2-5] Other components, such as sterols, fatty acids, and essential oil^[2,6-8] were also confirmed in P. villosa (Thunb.) Juss.

As part of our search for new natural compounds with anti-tumor activity, we carried out phytochemical investigations on the leaves of *P. villosa* (Thunb.) Juss., collected in China. A new biphenyl neolignan and four known compounds [Figure 1] were isolated from the plant. The present study reports the isolation and structural elucidation of these components.

MATERIALS AND METHODS

General experimental procedures

Optical rotations were measured on a Perkin-Elmer digital polarimeter (USA). nuclear magnetic resonance (NMR) spectra were recorded

with tetramethylsilane as internal standard on a Bruck AVANCE 400 FT-NMR spectrometer. Electrospray ionization/mass spectrometry was measured on an Agilent 1100 LC/MSD Trap-SL spectrometer (USA). Column chromatography was performed on silica gel (200–300 mesh) (Marine Chemical Factory, Qingdao, China) and octa decylsilyl silicion (ODS) (ultimate XB-C18, 40–70 μm , Welch Materials, Inc., USA). High-performance liquid chromatography (HPLC) separation was performed with an Agilent 1100 chromatograph apparatus using an ODS column (ultimate XB-C18, 10×250 , 5 μm , Welch Materials, Inc., USA) and detected with an ultraviolet detector.

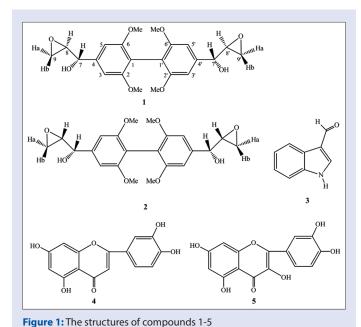
Materials and chemicals

The *P. villosa* Juss., leaves were purchased from Hebei Qixin Traditional Chinese Medicine Pellets Co., Ltd., P.R. China, and identified by Prof. Kang Tingguo, College of Pharmacy, Liaoning University of Traditional Chinese Medicine. A voucher specimen (NO. PVJ20130821) has been deposited at the Pharmacognosy Laboratory, Harbin University of

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Commerce. Organic solvents (analytical grade or HPLC grade) for the experiment were purchased from Kermel Chemical Co., (Tianjin, China).

Extraction and isolation

The air-dried leaves of P. villosa Juss., (15 kg) were extracted three times with 70% EtOH under reflux. Evaporation of the solvent under reduced pressure gave a condensed extract (2.8 kg), which was suspended in H₂O and then partitioned successively with light petroleum, dichloromethane, and n-BuOH. The dichloromethane portion (110 g) was then subjected to normal phase silica gel column chromatography and eluted with a gradient of CH₂Cl₂-MeOH (100:0 → 0:100, v/v) to give ten fractions. Fraction 2 (CH₂Cl₂-MeOH, 80:1, v/v) was subjected to further separation using ODS silica gel column chromatography eluted with 20%, 36%, 44%, 60%, and 80% MeOH in water, respectively. Subfraction 2.2 (36% methanol elution) was purified by semi-preparative HPLC (MeCN-H₂O, wavelength 280 nm) to yield compounds 1 (16.5 mg) and 2 (21.3 mg). Subfraction 2.3 (44% methanol elution) was purified by semi-preparative HPLC (MeCN-H₂O, wavelength 280 nm) to give compound 3 (10.4 mg). The n-BuOH portion (280 g) was subjected to normal phase silica gel column chromatography and eluted with a gradient of CH₂Cl₂-MeOH (100:0 \rightarrow 0:100, v/v) to give eight fractions. Fraction 4 was subjected to further separation using repeated ODS silica gel column and HPLC to give compounds 4 (25.0 mg) and 5 (18.6 mg).

RESULTS AND DISCUSSION

Compound 1 was obtained as a colorless needle crystal (CHCl $_3$), with the molecular formula $C_{22}H_{26}O_8$ as determined by the high-resolution electrospray ionization mass spectrometry (HRESIMS) at m/z 441.1518 (M + Na) $^+$, indicating ten degrees of unsaturation. The 1 H-NMR spectrum of compound 1 [Table 1] displayed four aromatic proton signals at $\delta_{\rm H}$ 6.59 (4H, s, H-3, H-3′, H-5, H-5′), which were assigned to two 1,2,4,6-tetrasubstituted benzene rings. In addition, the 1 H-NMR spectrum also revealed four oxymethine protons at $\delta_{\rm H}$ 4.74 (2H, d, J = 4.0 Hz, H-7, H-7′) and 3.10 (2H, m, H-8, H-8′), two methylene protons at $\delta_{\rm H}$ 4.28 (2H, m, H-9a, H-9′a) and 3.93 (2H, m, H-9b, H-9′b), four methoxyl groups at $\delta_{\rm H}$ 3.90 (12H, s). The 13 C NMR spectrum displayed 22 carbon signals, including 12 aromatic

Table 1: ¹H-NMR (300 MHz) and ¹³C-NMR (100 MHz) spectral data of compounds 1 and 2 in CDCl,

Number	1		2	
	$\boldsymbol{\delta}_{H}$	δ _c	$\boldsymbol{\delta}_{H}$	δ _c
1,1'		132.1		132.0
2,2'		147.2		147.1
3,3'	6.59 (s)	102.8	6.57 (s)	102.8
4,4'		134.34		134.3
5,5'	6.59 (s)	102.8	6.57 (s)	102.7
6,6'		147.2		147.1
7,7'	4.74 (d, 4.0)	86.1	4.78 (d, 7.0)	86.0
8,8'	3.10 (m)	54.4	3.09 (m)	54.3
9a, 9'a	4.28 (m)	71.8	3.89 (m)	71.8
9b, 9'b	3.93 (m)		4.26 (m)	
-OH	5.52 (s)		5.54 (s)	
-OCH ₃	3.90 (s)	56.4	3.90 (s)	56.3

Coupling constants (J) in Hz are given in parentheses; chemical shift values are expressed in ppm. NMR: Nuclear magnetic resonance

carbons, four oxygen-substituted methine ($\delta_{\rm C}$ 54.4, 54.4, 86.1, 86.1), two oxygen-substituted methylene ($\delta_{\rm C}$ 71.8, 71.8), and four methoxyl carbons ($\delta_{\rm C}$ 56.4), All of the above spectroscopic data were similar to those of 2,6,2′,6′-tetramethoxy-4,4′-bis (1,2-cis-2,3-epoxy-1-hydroxypropyl) biphenyl (2) except for difference of coupling constants at H-7, 7′ (J=4.0 Hz). In the NOESY experiment [Figure 2] on 1, correlations between H-3/H-7 or H-5/H-7, and a strong cross-peak between H-8/H_a-9 suggested the β -configuration for H-7 and H_b-9 and an α -configuration for OH-7. The relative configuration could be proposed as 7,8-trans-8,9a-cis. Thus, the structure of one was determined as 2,6,2′,6′-tetramethoxy-4,4′-bis (1,2-trans-2,3-epoxy-1-hydroxypropyl) biphenyl.

Compounds 2–5 were identified as 2,6,2',6'-tetramethoxy-4,4'-bis (2,3-epoxy-1-hydroxypropyl) biphenyl (2), 1H-indole-3-carbaldehyde (3), luteolin (4), and quercetin (5) on the basis of its spectroscopic data. [9-11]

CONCLUSION

One new stereoisomer of biphenyl neolignan (1) with four known compounds 2–5 was isolated from the leaves of *P. villosa* Juss. The structures of all the compounds were elucidated on the basis of spectroscopic analysis and comparison with literature data.

Compounds

2,6,2′,6′-Tetramethoxy-4,4′-bis (1,2-*trans*-2,3-epoxy-1- hydroxypropyl) biphenyl (1). Colorless needle crystal. HRESIMS m/z 441.1518 (M + Na)+ (calculated for C $_{22}$ H $_{26}$ O $_{8}$ Na, 441.1520). ¹H-NMR (CDCl $_{3}$, 300 MHz): $\delta_{\rm H}$: 3.10 (2H, m, H-8, H-8′), 4.28 (2H, m, H-9a, H-9′a), 3.90 (12H, s, OMe-2, OMe-2′, OMe-6, OMe-6′), 3.93 (2H, m, H-9b, H-9′b), 4.74 (2H, d, J = 4.0 Hz, H-7, H-7′), 5.52 (2H, brs, OH-7, OH-7′), 6.59 (4H, s, H-3, H-3′, H-5′, H-5′). ¹³C-NMR $\delta_{\rm C}$: 54.4 (2C, C-8, C-8′), 56.4 (4C, OMe-2, OMe-2′, OMe-6, OMe-6′), 71.8 (2C, C-9, C-9′), 86.1 (2C, C-7′, C-7′), 102.8 (4C, C-3, C-3′, C-5, C-5′), 132.1 (2C, C-1, C-1′), 134.3 (2C, C-4, C-4′), 147.2 (4C, C-2, C-2′, C-6, C-6′).

2,6,2′,6′-Tetramethoxy-4,4′-bis (1,2-*cis*-2,3-epoxy-1-hydroxypropyl) biphenyl (2). Colorless needle crystal. ¹H-NMR (CDCl₃, 300 MHz): $\delta_{\rm H}$: 3.09 (2H, m, H-8, H-8′), 3.89 (2H, m, H-9a, H-9′a), 3.90 (12H, s, OMe-2, OMe-2′, OMe-6, OMe-6′), 4.26 (2H, m, H-9b, H-9′b), 4.78 (2H, d, J = 7.0 Hz, H-7, H-7′), 5.54 (2H, brs, OH-7, OH-7′) 6.57 (4H, s, H-3, H-3′, H-5, H-5′). ¹³C-MNR $\delta_{\rm C}$: 54.3 (2C, C-8, C-8′), 56.3 (4C, OMe-2, OMe-2′, OMe-6, OMe-6′), 71.8 (2C, C-9, C-9′), 86.0 (2C, C-7, C-7′),

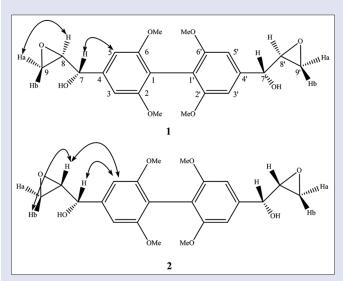


Figure 2: Key NOESY correlations for 1 and 2

102.7 (4C, C-3, C-3', C-5, C-5'), 132.0 (2C, C-1, C-1'), 134.3 (2C, C-4, C-4'), 147.1 (4C, C-2, C-2', C-6, C-6').

1H-indole-3-carbaldehyde (3). Pale yellow needle crystal. 1 H-NMR (dimethyl sulfoxide [DMSO], 400 MHz) $\delta_{\rm H}$: 7.24 (2H, m, 5,6-H), 7.52 (1H, d, J=8.0 Hz, 7-H), 8.10 (1H, s, 2-H), 8.29 (1H, d, J=7.0 Hz, 4-H). 13 C-NMR $\delta_{\rm C}$: 185.4 (C-10), 138.9 (C-8), 137.5 (C-2), 124.6 (C-9), 123.9 (C-5), 122.6 (C-6), 121.3 (C-7), 118.64 (C-3), 112.6 (C-4).

Luteolin (4). Pale yellow powder. $^{1}\text{H-NMR}$ (DMSO, 400 MHz) δ_{H} : 13.02 (1H, brs, 5-OH), 7.45 (1H, dd, J=8.5, 2.5 Hz, 6'-H), 7.40 (1H, d, J=2.5 Hz, 2'-H), 6.91 (1H, d, J=8.5 Hz, 5'-H), 6.71 (1H, s, 3-H), 6.47 (1H, d, J=2.0 Hz, 6-H), 6.59 (1H, d, J=1.6 Hz, H-8). $^{13}\text{C-NMR}$ δ_{C} : 163.8 (C-2), 103.3 (C-3), 181.4 (C-4), 161.4 (C-5), 99.0 (C-6), 164.9 (C-7), 93.9 (C-8), 157.3 (C-9), 103.9 (C-10), 118.8 (C-1'), 113.1 (C-2'), 145.9 (C-3'), 149.9 (C-4'), 116.0 (C-5'), 121.7 (C-6').

Quercetin (5). Pale yellow powder. ^1H -NMR (DMSO, 400 MHz) δ_{H} : 6.20 (1H, d, J = 2.0 Hz, 6-H), 6.38 (1H, d, J = 2.0 Hz, 8-H), 6.86 (1H, d, J = 8.4 Hz, 5′-H), 7.56 (1H, dd, J = 8.4, 2.0 Hz, 6′-H), 7.65 (1H, d, J = 2.0 Hz, 2′-H), 9.32, 9.38, 9.62, 10.79, 12.49 (1H, s,-OH × 5). ^{13}C -NMR δ_{C} : 146.9 (C-2), 122.1 (C-3), 175.9 (C-4), 156.3 (C-5), 98.3 (C-6), 164.0 (C-7), 93.4 (C-8), 160.9 (C-9), 103.3 (C-10), 123.3 (C-1′), 115.8 (C-2′), 148.0 (C-3′), 145.2 (C-4′), 115.2 (C-5′), 120.2 (C-6′).

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Conflicts of interest

There are no conflicts of interest.

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